Laboratory for Male Reproductive Research and Testing

Direct and Indirect Immunobead (IMB)

IMB is measured in the laboratory on fresh ejaculates. Please prepare the sample as follows:

**Indirect Immunobead Test**

1.) Please ask your patient to abstain from sexual activity 3 days before he collects his sample. Semen kept at 37°C should liquefy 30 minutes after collection. Measure the volume on the graduated cylinder provided and perform a routine semen analysis. On the attached form, please record:
   - Volume
   - Sperm concentration (million/mL)
   - Percent of moving (% motility)
   - Average forward progression (0-4)
   - Total number of motile sperm in the ejaculate (multiply semen volume by the sperm concentration by the % motility)
   - Time of analysis

2.) Mail by FedEx (overnight delivery). Place two ice packs on either side of bottle and place in a styrofoam box.

**Direct Immunobead Test (live sperm)**

Controlling the temperature of the semen during shipping is very important. This kit includes a plastic bottle, which will keep tubes of semen submerged in cold water during transit. The volume of water will prevent the samples from accidentally freezing or becoming too warm. This will enable us to more accurately control sample temperature and ensure motility is maintained.

1.) Thaw a tube of test yolk buffer (TYB) cryopreservative 30 minutes before you wish to use it. If you are supplied with additional tubes of TYB, they can be stored indefinitely in the freezer.

2.) Please ask your patient to abstain from sexual activity 3 days before he collects his sample. Semen kept at 37°C should liquefy 30 minutes after collection. Measure the volume on the graduated cylinder provided and perform a routine semen analysis. On the attached form, please record:
   - Volume
   - Sperm concentration (million/mL)
   - Percent of moving (% motility)
   - Average forward progression (0-4)
   - Total number of motile sperm in the ejaculate (multiply semen volume by the sperm concentration by the % motility)
   - Time of analysis
3.) Add equal volume of room temperature TYB to all of the remaining semen and mix with the aid of a Pasteur pipette. Hyperviscous samples may require drawing the mixture in and out of the pipette 10 or more times to achieve adequate mixing. Label the tube with the patient’s name using an indelible marker. An adequate seal is performed when the cap is on tight.

4.) It is important to cool this mixture gradually. This can easily be accomplished by filling the plastic bottle provided with room temperature water. Submerge the tube containing the mixture in the water and the bottle tightly. Place it in the refrigerator (not freezer) and record the time on the form. No more than five tubes can fit inside the shipping bottle. **No more than 1 hour should elapse from the time the sample was collected and the cooling process.**

5.) Refrigerate the mixture for at least 4 hours, then “sandwich” the bottle between the two pre-frozen ice packs provided. Place the bottle and ice packs in the Styrofoam shipping box so that the contents will not shift at liberty. A space can be molded to more conveniently accept the sample bottle by freezing the packs earlier with the empty sample bottle between them.

6.) If any additional supplies are needed, simply include a note describing your specific needs, and they will arrive free of charge on the following Friday via FedEx.

Direct Immunobead tests are done by appointment only so that we can anticipate the number of assays to be run. This will also ensure that holidays and vacations will not conflict with processing the sample. **Direct** Immunobead can only be done on a patient with a sperm concentration of at least 60 million/mL and a motility of 50%.

Payment for each IMB assay should accompany the samples. **Make checks payable to the Laboratory for Male Reproductive Research and Testing (LMRRT).** Please make sure that the patient’s name is on the check. We do not accept insurance assignments, file insurance claims, bill neither patient, nor contract with insurance companies. We only complete necessary information of the HICFA form and forward it to you – if payment is received. **If payment is not received,** the lab will invoice the requesting/referring physician or clinic.

Please call the lab at (713) 798-7265 to schedule the test.

♦ **Shipping containers with supplies can be mailed upon request.**

♦ **Lab address:**
  LMRRT
  Baylor College of Medicine
  One Baylor Plaza, N730; MS380
  Houston, TX 77030
  Telephone: (713) 798-7265
  Fax: (713) 798-1891